

DEISOVALERYLBLASTMYCIN PRODUCED BY *STREPTOMYCES* SP.\*

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A new antifungal antibiotic was isolated from the fermentation broth of *Streptomyces* sp. 5140-A<sub>1</sub>. Degradation studies of the crystalline antibiotic, m.p. 186~188°C, C<sub>21</sub>H<sub>28</sub>O<sub>8</sub>N<sub>2</sub>, suggested its structure to be deisovaleryl-blastmycin. It exhibited antimicrobial activity against *Piricularia oryzae* and less toxicity against killifish than antimycin A—blastmycin antibiotics.

During screening of new antibiotics, a strain of *Streptomyces* sp. 5140-A<sub>1</sub>, isolated from a soil sample collected at Moiwa Forest, Sapporo-city, Japan, was found to produce a new antifungal antibiotic, which was closely related to the antimycin A—blastmycin group of antibiotics<sup>1-5</sup>). Its structure was finally determined as deisovaleryl-blastmycin (II). In this report, fermentation, purification procedures, and chemical and biological properties are discussed.

#### Fermentation

*Streptomyces* sp. 5140-A<sub>1</sub> was cultivated in the medium containing starch 2.5%, soybean meal 1.5%, dry yeast 0.2%, ammonium sulfate 0.2%, sodium chloride 0.5% and calcium carbonate 0.4% at pH 6.2 (before sterilization).

Antibiotic production rose to a maximum 24~28 hours after inoculation, and differential bioassay using *Piricularia oryzae* as a test organism showed 10~20 mcg/ml of new antibiotic accumulated together with blastmycin in the fermentation broth.

#### Isolation Procedure

Cultured broth of the streptomyces was harvested after 24 hours fermentation and the mycelial cake was extracted with 60% aqueous acetone. After removal of the acetone, the aqueous phase was extracted with ethyl acetate. The organic layer, combined with the ethyl acetate extract of the filtered beer, was washed with water, dried over sodium sulfate and concentrated *in vacuo* to a brownish solution, which was passed through a silica gel column.

The colorless active filtrate was concentrated and applied to a silicic acid column, which was washed with benzene, then developed carefully with benzene - ethyl acetate, whereby two active fractions were separated. From the first fraction was isolated crystalline blastmycin. Later fractions were concentrated and diluted with hexane to give colorless crystals, which were recrystallized from ethyl acetate - hexane or aqueous methanol.

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In another procedure, blastmycin was removed with benzene from the fermentation beer or mycelial extract, then the deacyl compound was extracted from the residual liquor with butanol, and purified by silicic acid column chromatography. This method was applied for differential assay of fermentation products.

#### Physicochemical Properties

The colorless crystalline compound had m.p. 186~188°C and  $[\alpha]_D^{25} +36^\circ$  (*c* 1.0, methanol). It gave positive color reactions with ferric chloride and LEMIEUX's reagent, but negative with ninhydrin, FEHLING and silver nitrate reagents. It is soluble in alcohols, ethyl acetate and acetone, slightly soluble in benzene, ether and chloroform, and almost insoluble in petroleum ether, hexane and water. Elemental analysis and molecular weight determination by mass spectrometry suggested  $C_{21}H_{28}O_8N_2$  for its molecular formula.

Calcd. for  $C_{21}H_{28}O_8N_2$ :  
 C 57.79, H 6.47, O 29.33, N 6.42  
 MW 436.45  
 Found; C 57.99, H 6.63, O 28.88, N 6.42  
 MW 436 ( $M^+$ )

Fig. 1. UV spectra of deisovaleryl-blastmycin

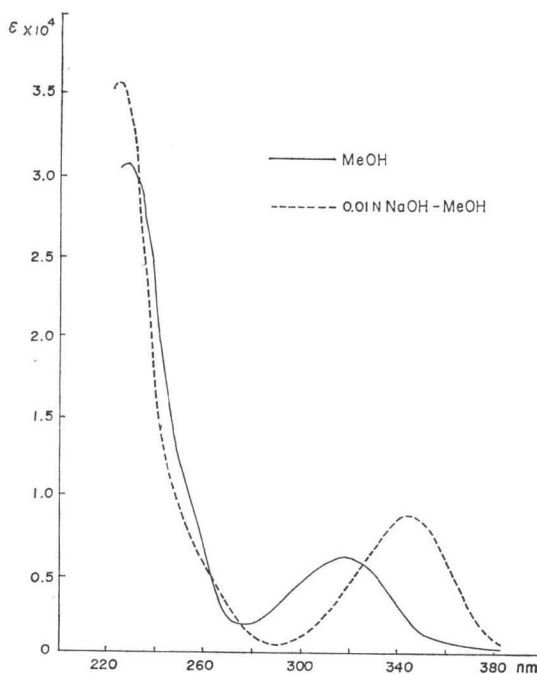


Fig. 2. IR spectrum of deisovaleryl-blastmycin ( $CHCl_3$ )

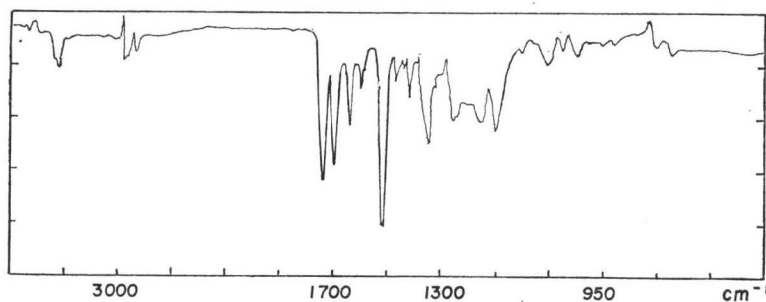
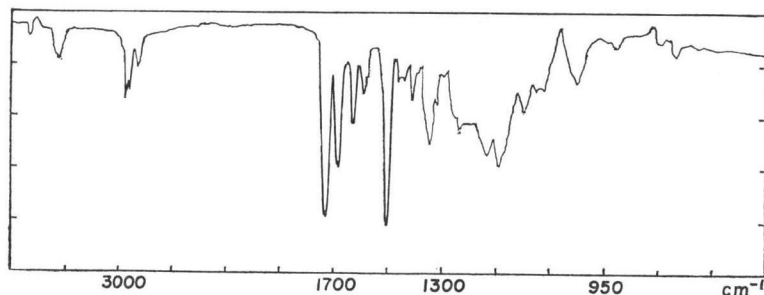


Fig. 3. IR spectrum of blastmycin ( $CHCl_3$ )



The UV spectra of the crystalline compound (Fig. 1) show maxima at  $\lambda_{\text{max}}$  226 nm ( $\epsilon$  30,400), 320 (6,000) in methanol and *N*/100 HCl-MeOH and at  $\lambda_{\text{max}}$  225 nm ( $\epsilon$  35,300), 345 (8,500) in *N*/100 NaOH-MeOH.

These maxima are essentially the same as those of antimycin A or blastmycin, except for their molecular extinction coefficients. The differences between the crystalline antibiotic and these antibiotics are shown in their IR spectra (Figs. 2 and 3) and in their thin-layer chromatograms. The crystalline antibiotic had *R<sub>f</sub>* values of 0.25 and 0.16 in benzene-ethyl acetate (3 : 2) and chloroform-methanol (20 : 1) system respectively, whereas blastmycin had *R<sub>f</sub>* 0.70 and 0.90.

### Chemical Structure

The new antibiotic (II) was considered to be a compound similar to antimycin A or blastmycin (I) from the data mentioned above. Similarity of UV spectra, and the detection of L-threonine from the hydrolysate of II (5.7*N* HCl, 120°C, 12 hours) showed the same amino acid composition as I. Formation of  $\alpha$ -butyllevulinic acid (VII) by vigorous alkaline hydrolysis (3 *N* NaOH, 100°C, 1 hour), but no volatile acid (VI) suggested the absence of the acyl group at R' position, as in formula (II).

When I and II were acylated in isovaleric anhydride-pyridine solution, acyl-I and acyl-II showed the same mobility on TLC.

Mild alkaline hydrolysis of antimycin A or blastmycin (I) (1 *N* NaOH, room temperature, few minutes) gave blastmycic acid (IV) and antimycinone or blastmycinone (III), which are mixtures of alkyl

Fig. 4. Degradation scheme of blastmycin and de-isovaleryl-blastmycin

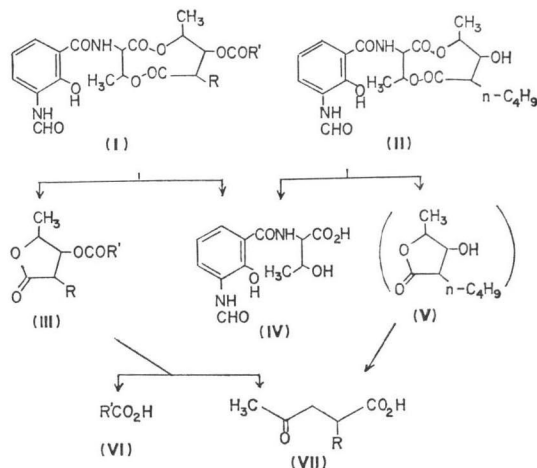


Fig. 5. Gas chromatograms of blastmycinones.

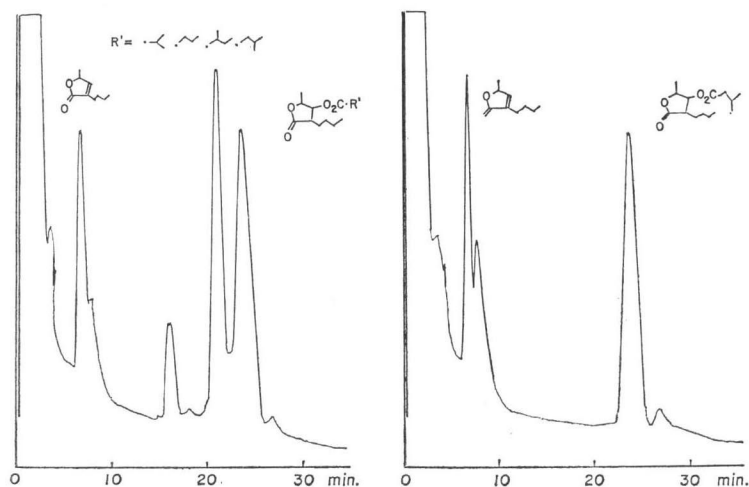


Table 1. Antifungal activity of deisovalerylblastmycin.

Fungi	M. I. C. (mcg/ml)
<i>Alternaria kikuchiana</i> IAM-5005	25
<i>Aspergillus oryzae</i>	>100
<i>Botrytis cinerea</i> IAM-5127	>100
<i>B. fabae</i> IAM-5125	>100
<i>Cladosporium fulvum</i> NIAS	50
<i>Fusarium lini</i> NIAS	>100
<i>Gibberella fujikuroi</i> NIAS	>100
<i>G. saubineti</i> NIAS	50
<i>Gloeosporium kaki</i> NIAS	50
<i>Glomerella cingulata</i> IAM-8050	>100
<i>G. lagenarium</i> NIAS	>100
<i>Helminthosporium sesamum</i> NIAS	12.5
<i>H. sigmoideum</i>	100
<i>Macrosporium bataticola</i> IAM-5014	>100
<i>Mucor ramannianus</i> IAM-6128	>100
<i>Ophiobolus miyabeanus</i> NIAS	25
<i>Penicillium chrysogenum</i> Q 176	25
<i>Piricularia oryzae</i> NIAS	0.8
<i>Trichophyton mentagraphytes</i> NIHJ-640	25
<i>Candida albicans</i> IAM-4905	>100
<i>C. utilis</i> IAM-4215	3.125
<i>Cryptococcus neoformans</i> IAM-4514	>100
<i>Rhodotorula glutinis</i> IAM-4757	50
<i>Saccharomyces cerevisiae</i> NIHJ-F-130	>100

Table 2. Preventive effect of deisovalerylblastmycin against rice-blast disease.

DV-BM (ppm)	Blasticidin S (ppm)	Spots/leaf	Effectivity (%)	Phyto-toxicity
400	0	4.0	86.1	—
200	0	17.5	39.2	—
100	0	16.7	42.0	—
0	10	1.1	96.2	—
0	0	28.8	0.0	—

on pot test in a green house, and was 86.1% effective at 400 ppm concentration.

### Discussion

Deisovalerylblastmycin (II) was isolated from the fermentation broth of *Streptomyces* sp. 5140-A<sub>1</sub> in an early stage of the fermentation.

Recently, SINGH *et al.*<sup>12)</sup> reported the enzymatic preparation of deacylantimycin A by hog kidney deacylase. In the present case, the deacyl compound was isolated from the same *Streptomyces* which produces blastmycin (I, R = *n*-C<sub>4</sub>H<sub>9</sub>- and R' = (CH<sub>3</sub>)<sub>2</sub>CHCH<sub>2</sub>-) and with regard to the fermentation time of this strain, the deacyl compound may be a biosynthetic precursor of blastmycin.

residues at R and R' and which were separated on gas chromatograms (20% DEGS, Fig. 5)<sup>6)</sup>. Under these conditions, the blastmycin mixture was observed to be a mixture of R' = (CH<sub>3</sub>)<sub>2</sub>-CH-, *n*-C<sub>8</sub>H<sub>7</sub>-, C<sub>2</sub>H<sub>5</sub>(CH<sub>3</sub>)CH- and (CH<sub>3</sub>)<sub>2</sub>CH-CH<sub>2</sub>- with R = *n*-C<sub>4</sub>H<sub>9</sub>-, and our antimycin mixture so far determined was a mixture of R = *n*-C<sub>4</sub>H<sub>9</sub>-, *n*-C<sub>6</sub>H<sub>13</sub>- and R' = (CH<sub>3</sub>)<sub>2</sub>CH-, *n*-C<sub>8</sub>H<sub>7</sub>-, C<sub>2</sub>H<sub>5</sub>(CH<sub>3</sub>)CH-, (CH<sub>3</sub>)<sub>2</sub>CHCH<sub>2</sub>-.

Isovalerylated II, on mild alkaline treatment, gave only a peak corresponding to R = *n*-C<sub>4</sub>H<sub>9</sub>- and R' = (CH<sub>3</sub>)<sub>2</sub>CHCH<sub>2</sub>- moiety on gas chromatography. For these reasons, structure II was assigned to the original antibiotic, named deisovalerylblastmycin.

As antimycin A components, more than twenty compounds have been detected with combinations of R (C<sub>2</sub>~C<sub>8</sub>) and R' (C<sub>1</sub>~C<sub>7</sub>) side chains<sup>6-11)</sup> and the present antibiotic is characterized by no acyl group at R' position.

### Biological Properties

The antimicrobial activity of deisovalerylblastmycin is shown in Table 1. It is primarily active against *Piricularia oryzae* and *Candida utilis*, and weakly active against some fungi, but not active against species of bacteria. The LD<sub>50</sub> of deisovalerylblastmycin was 15 mg/kg (i.v., dd mice) and 25 mg/kg (i. p.).

Toxicity against fish was 0.5 mcg/ml (killifish), 1/1,000 as toxic as antimycin A or blastmycin mixtures.

The preventive effect of deisovalerylblastmycin against rice-blast disease was determined

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